

# Evaluating Asian Chestnut Gall Wasp and Developing an Integrated Management Strategy

---

Louise Labbate, M.S. Graduate Student

Dr. Deborah G. McCullough, Professor

MSU Dept. of Entomology & Dept. of Forestry

**Collaborators:** Dr. Dennis Fulbright, Dr. Carmen Medina Mora,  
Mario Mandujano, Erin Lizotte (MSU)  
Dr. Phillip Lewis (USDA APHIS)



# Asian chestnut gall wasp - *Dryocosmus kuriphilus* Yasumatsu (Hymenoptera: Cynipidae)

## Invasion History

- Native to China
- Japan 1941
- Korea 1961
- United States 1974
- Europe 2002



# Asian Chestnut Gall Wasp

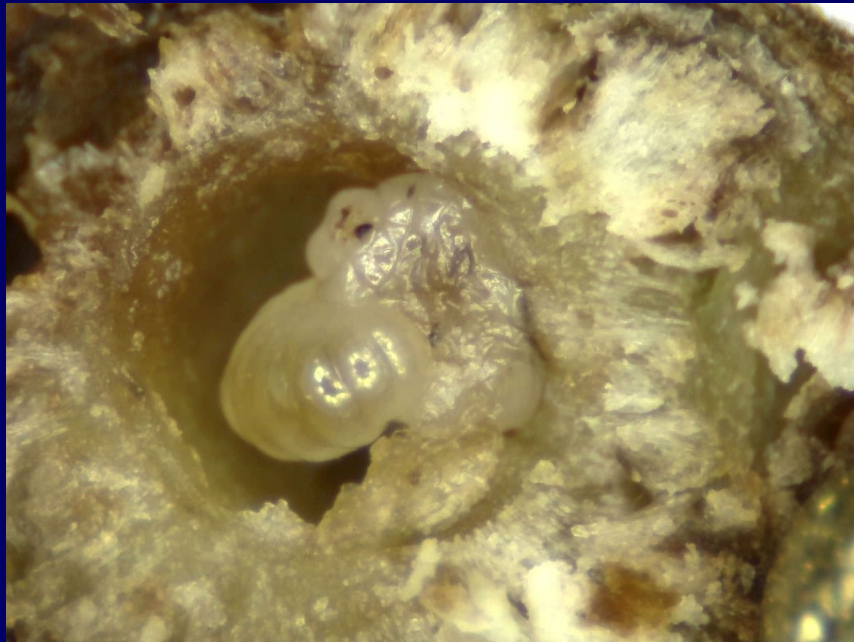
## Life Cycle

- Parthenogenic
- Univoltine
- Fall: Adults oviposit in buds
- Winter: 1<sup>st</sup> instar larvae overwinter in buds
- Spring: Larval feeding induces galls
- Summer: Adults emerge & lay eggs



## ACGW classical biocontrol with *Torymus sinensis*

- *Torymus sinensis* was first released in 1975 in Japan for ACGW biocontrol
- Phenologically synchronized with ACGW
  - Adults emerge in early spring as galls are forming
  - Lay 1 egg on an early instar ACGW larva (in gall)
  - *T. sinensis* larvae feed on ACGW larvae
  - Pupate in late winter; remain in galls all winter



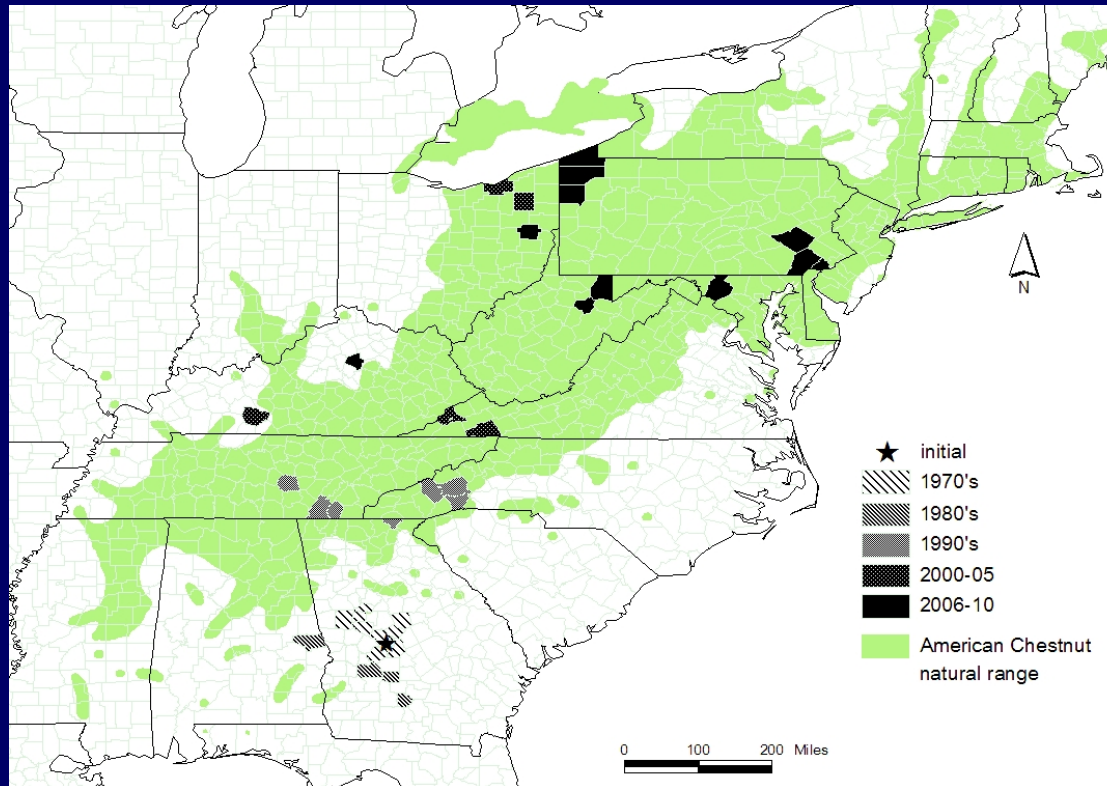
# Asian Chestnut Gall Wasp Spread

- Natural dispersal - flight
  - Short range
  - Wind-aided
  
- Artificial dispersal
  - Chestnut cuttings
  - Scion wood
  - Nursery trees



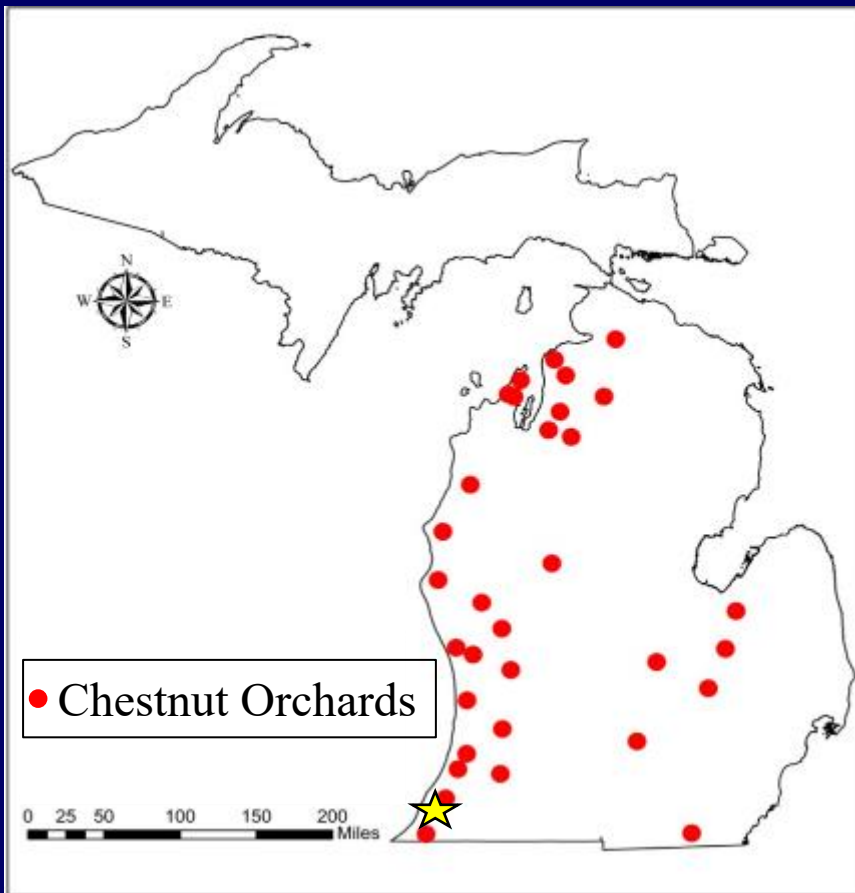
# ACGW in the United States

- First detected in Georgia in 1974 & was established in 11 other states by 2012
- Native parasitoids occasionally observed in galls
- *T. sinensis* parasitoid first released in Georgia in 1977



# ACGW in Michigan

- First Michigan detection was in 2015 in Berrien County
- By spring 2018, 10 chestnut orchards were infested, all within 45 miles of the first ACGW detection.



Locations in red represent 32 of at least 115 commercial chestnut orchards

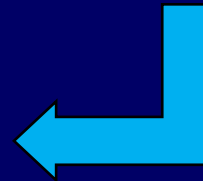
# Asian Chestnut Gall Wasp

## Damage Caused by ACGW

- Galls form on leaves & shoots
- Shoot growth & leaf area reduced
- **Apical galls** prevent flower & burr production



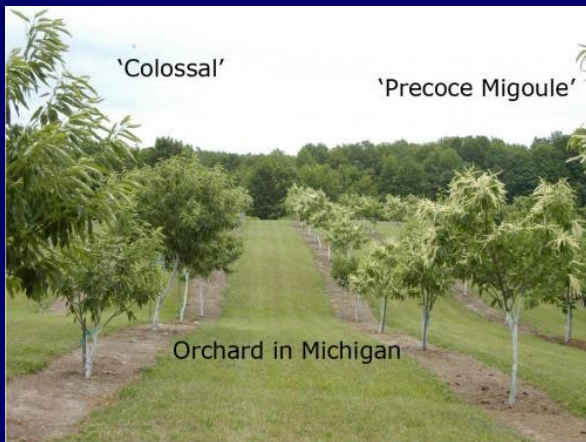
Yield Loss  
\$\$\$





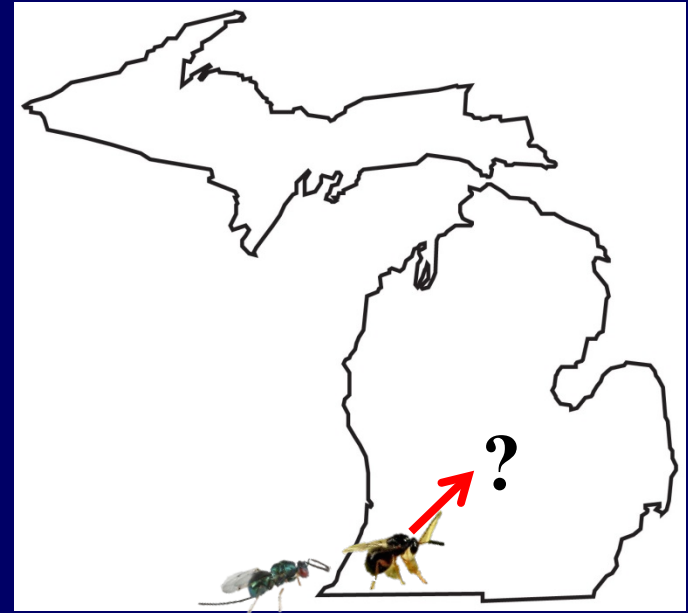
# Current ACGW Management Options

- Insecticide cover spray – effectiveness often limited; drift & non-target impacts can be problems
- Prune & destroy infested shoots - labor intensive, can result in major yield loss
- Resistant cultivars – some promise, ACGW adaptation; only relevant for new plantings
- Classical biocontrol with the *Torymus sinensis* parasitoid from China



# Why Study ACGW in Michigan?

- Harsh climate
- New invasion
  - ACGW spread
  - *T. sinensis* spread
  - Native parasitoids?
  - Invasion biology, spread, population dynamics



# Objectives

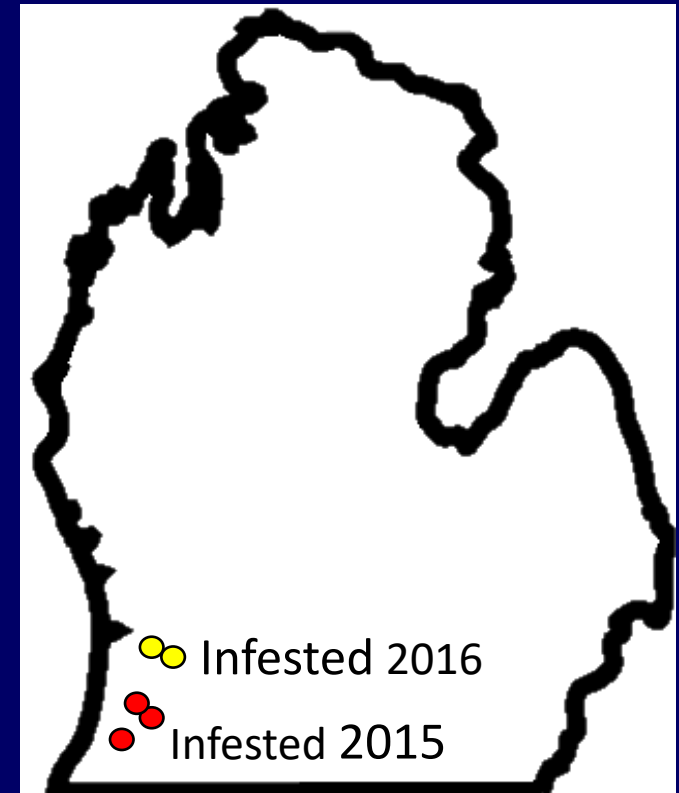
---

- Monitor phenology of chestnut trees & ACGW;  
Relate key stages to cumulative degree days (base 50 F)
- Quantify gall densities on selected chestnut cultivars & relate to shoot growth & burr production
- Evaluate parasitism: *Torymus sinensis* presence & rates
- Evaluate systemic insecticides: assess control of ACGW & persistence in selected chestnut tissues
- Assess spatial and temporal dynamics of ACGW & the *T. sinensis* parasitoid in commercial orchards in southwest Michigan.

# Study Sites - 2017

- 5 infested chestnut orchards
- Size ranged from 0.8 acres to 8 acres
- Sites located within 36 miles of each other
- Up to seven cultivars available for study

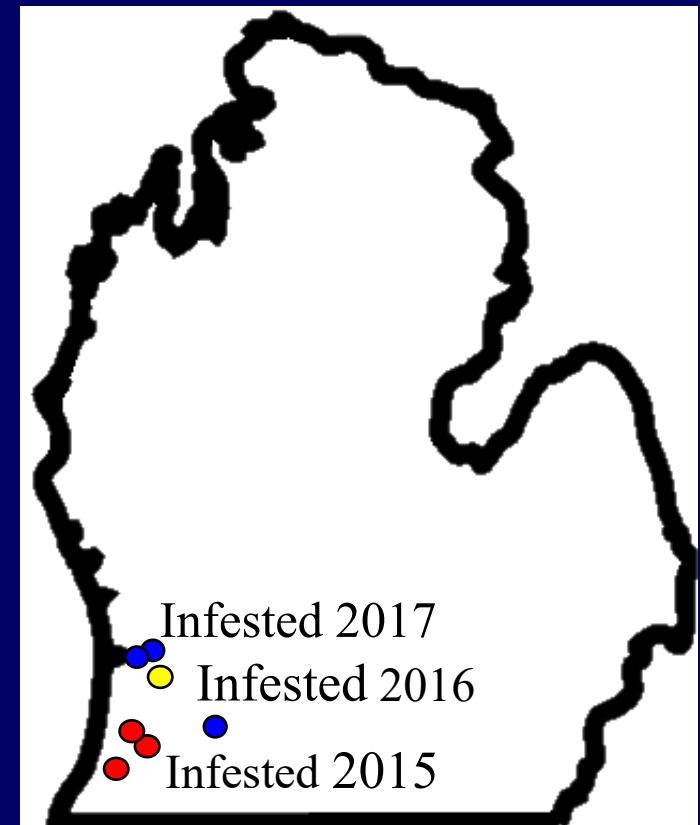
| Site | Species                       | Infested |
|------|-------------------------------|----------|
| 1    | Chinese, Euro x Japan, Korean | 2015     |
| 2    | Chinese                       | 2015     |
| 3    | Chinese, Euro x Japan         | 2015     |
| 4    | Chinese                       | 2016     |
| 5    | Chinese                       | 2016     |



# Study Sites - 2018

- 7 infested chestnut orchards
- Size ranged from 0.7 acres to 6.3 acres
- Sites located within 52 miles of each other
- Up to seven cultivars available for study

| Site | Species                       | Infested |
|------|-------------------------------|----------|
| 1    | Chinese, Euro x Japan, Korean | 2015 ●   |
| 2    | Chinese                       | 2015 ●   |
| 3    | Chinese, Euro x Japan         | 2015 ●   |
| 4    | Chinese                       | 2016 ●   |
| 5    | Chinese                       | 2017 ●   |
| 6    | Chinese, Euro x Japan         | 2017 ●   |
| 7    | Chinese                       | 2017 ●   |



# Chestnut Phenology

- Visually monitored chestnut trees to track developmental stages
  - Bud break
  - Catkin development
  - Pollen development
  - Burr production
- Degree days acquired from MSU Enviro-Weather station at SWMREC (all sites within 45 miles)



## Growing Degree Days Base 50 ° F

---

- Threshold (base temperature) is 50° F. Little or no insect development or activity occurs below 50 ° F.
- $DD_{50F}$  = maximum temperature minus the minimum temperature divided by 2, then subtract 50.
- MSU Enviro-weather stations across Michigan calculate cumulative  $DD_{50F}$  (Baskerville-Emin method).
- Degree days provide a means to track & anticipate development, regardless of calendar date.
- <https://enviroweather.msu.edu/>

# Chestnut Phenology & Cumulative Degree Days<sub>50F</sub>

| Chestnut Phenology                  | 2017 DD <sub>50F</sub> | 2018 DD <sub>50F</sub> |
|-------------------------------------|------------------------|------------------------|
| Bud Break                           | 273                    | 184                    |
| Leaves expanding                    | 354 to 615             | 238 to 529             |
| 1 <sup>st</sup> evidence of catkins | 354                    | 293                    |
| 1 <sup>st</sup> evidence of pollen  | 948                    | 905                    |
| Peak pollen                         | 1048                   | 1051                   |
| 1 <sup>st</sup> evidence of burrs   | 1368                   | 1138                   |
| Burrs mostly fallen                 | 2809                   | 3047                   |



- Early May: Bud break
- Late June: Peak pollen
- Early July: 1<sup>st</sup> burrs



## Potential Relevance

---

- Weather varies among years & regions. Degree days are more reliable than calendar dates.
- Relating developmental stages to degree days means information is applicable to other areas of the state.
- Comparing tree development to ACGW phenology can be useful for timing scouting or control activities.



# Gall Density – Methods 2017 & 2018

- Sites were visited every 2 weeks; mid May to mid July
- 2017: Sampled 2 shoots per tree from 9 to 18 trees per site in (613 shoots total)
- 2018: Sampled 1 shoot per tree from 4 to 40 trees per site (370 shoots total)
- Recorded gall location & density on collected shoots



# Gall Location

## ➤ Leaf galls

- Within leaf or petiole
- Minimal impact on trees



## ➤ Lateral galls

- Along branches
- May limit lateral shoot production

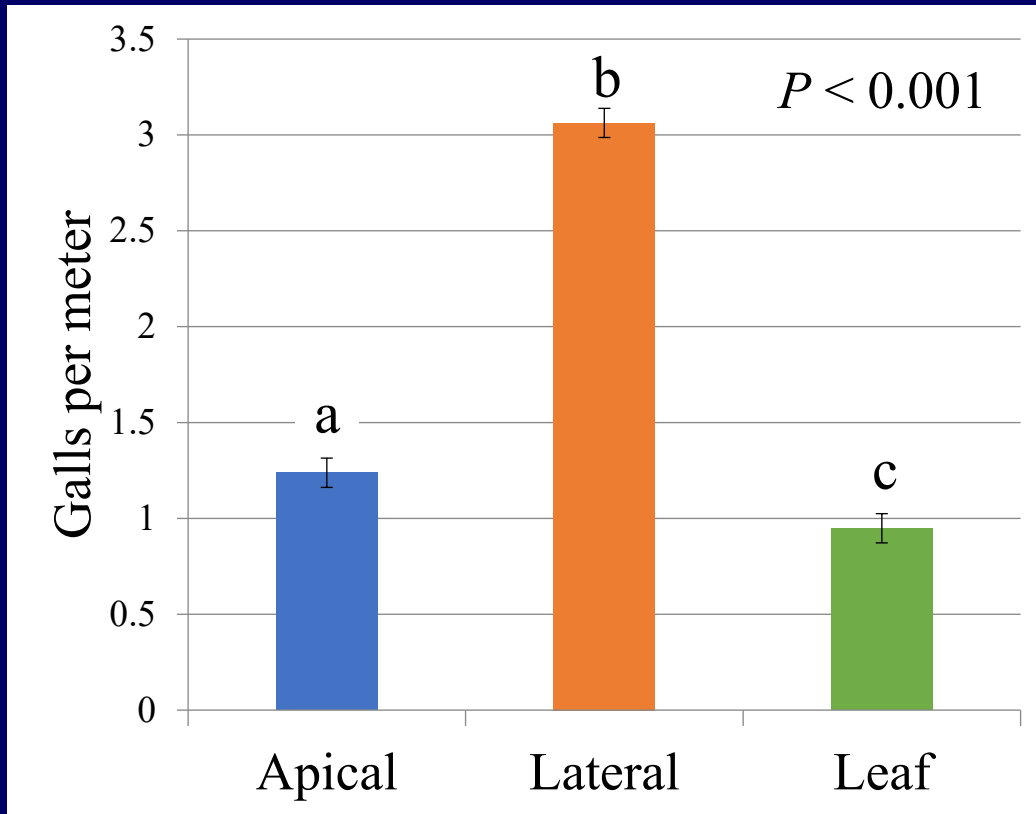


## ➤ Apical galls

- Ends of branches
- Potentially most damaging



# Average ( $\pm$ SE) density of current-year galls from 2017 & 2018 by gall location (n = 978 shoots)



Out of 6,488 total galls:

- 23% Apical
- 58% Lateral
- 19% Leaf

# Evaluating relative resistance or tolerance of two common cultivars to ACGW

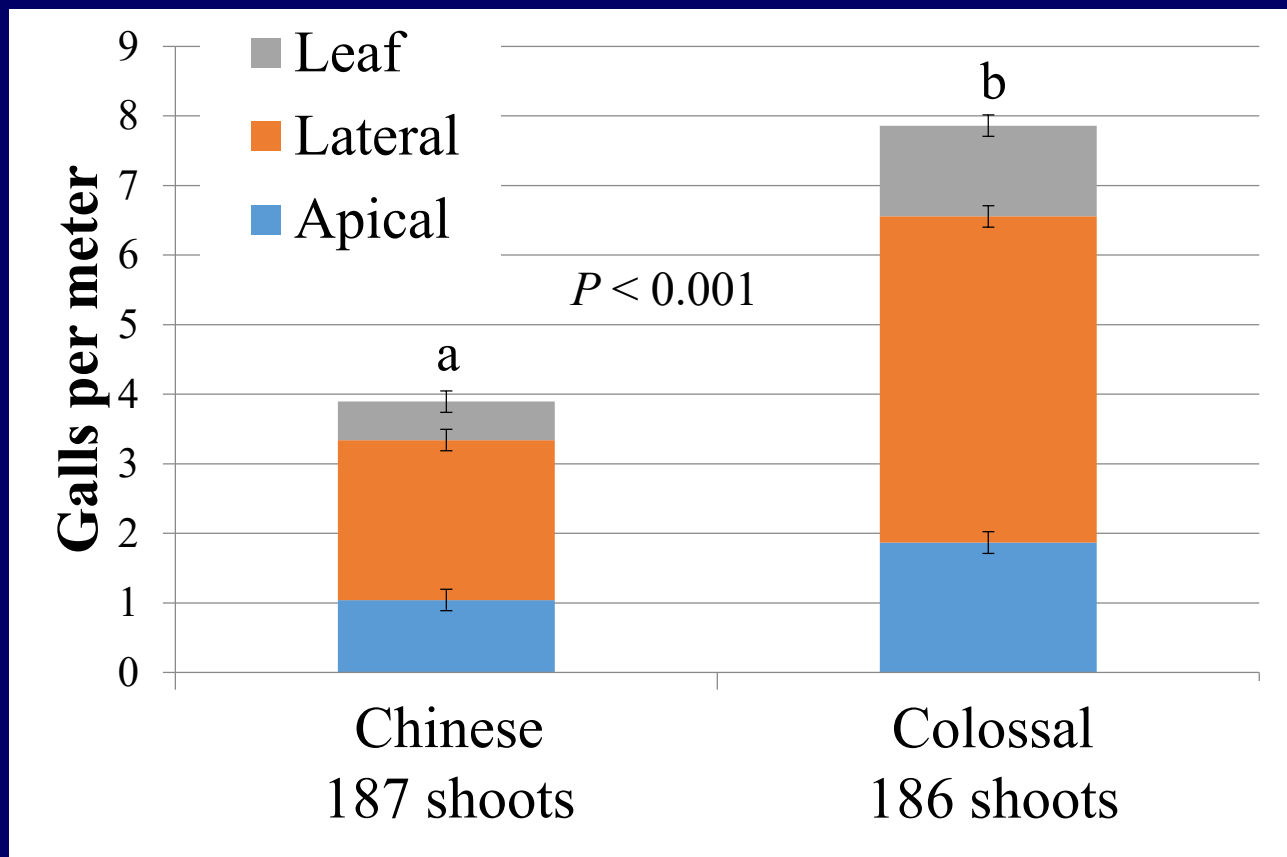
## Colossal Chestnut



## Chinese Chestnut



# Average ( $\pm$ SE) Density of Galls by Location on Colossal & Chinese Trees



Lateral Density:  
Different  
 $P < 0.001$

Leaf Density:  
Different  
 $P < 0.001$

Apical Density:  
Different  
 $P < 0.001$

➤ Total gall density is lower on Chinese trees

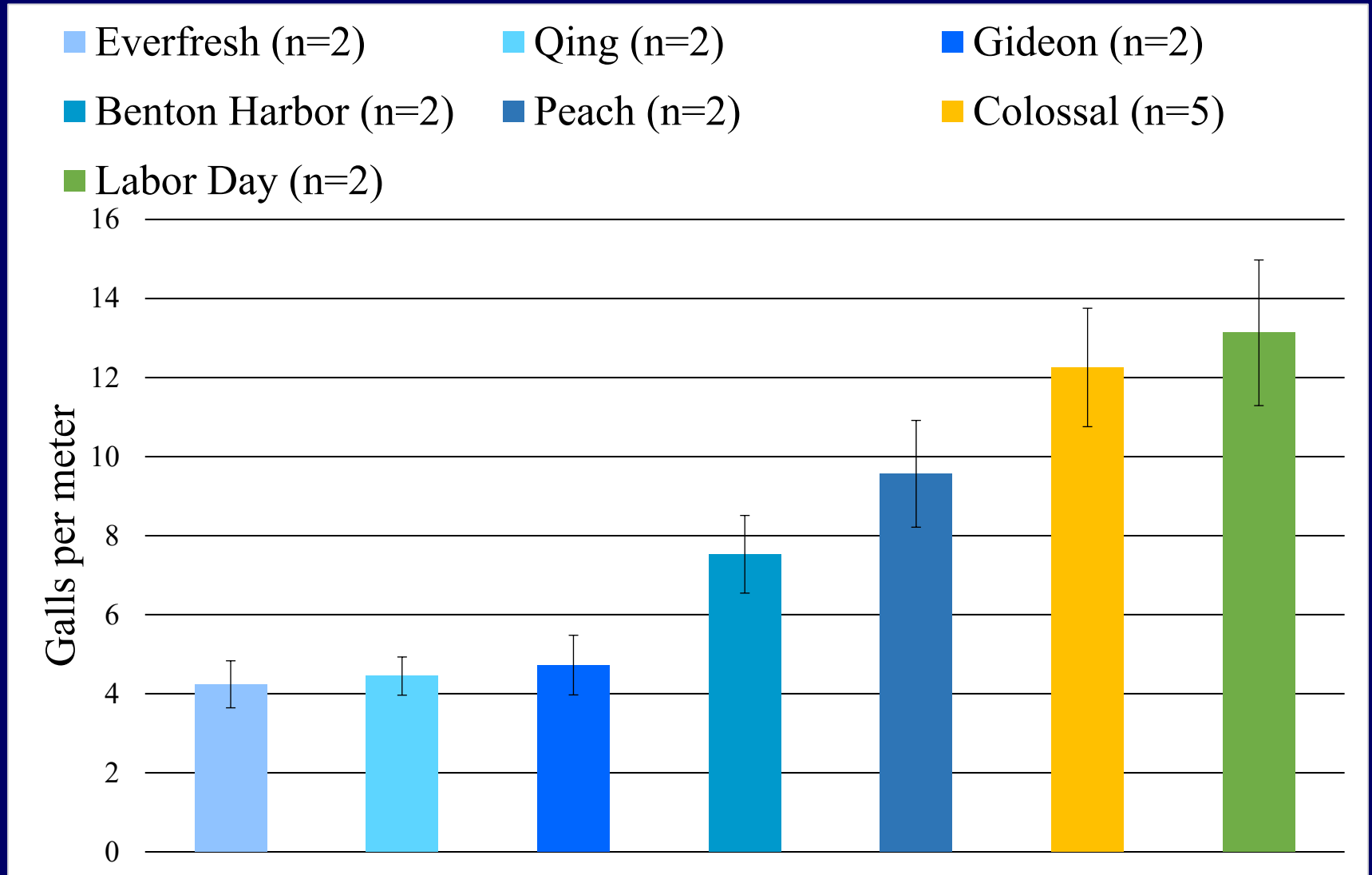
➤ Density of all gall types is lower on Chinese trees

# SWMREC Cultivar Study 2017 & 2018

- 3 species:
  - Chinese (*Castanea mollissima*)
  - European x Japanese (*C. sativa* x *C. crenata*)
  - Korean/Japanese (*C. crenata*)
- 42 cultivars planted – often represented by a single tree
  - Many newly planted or grafted trees
- 7 cultivars with 2-5 mature trees were sampled

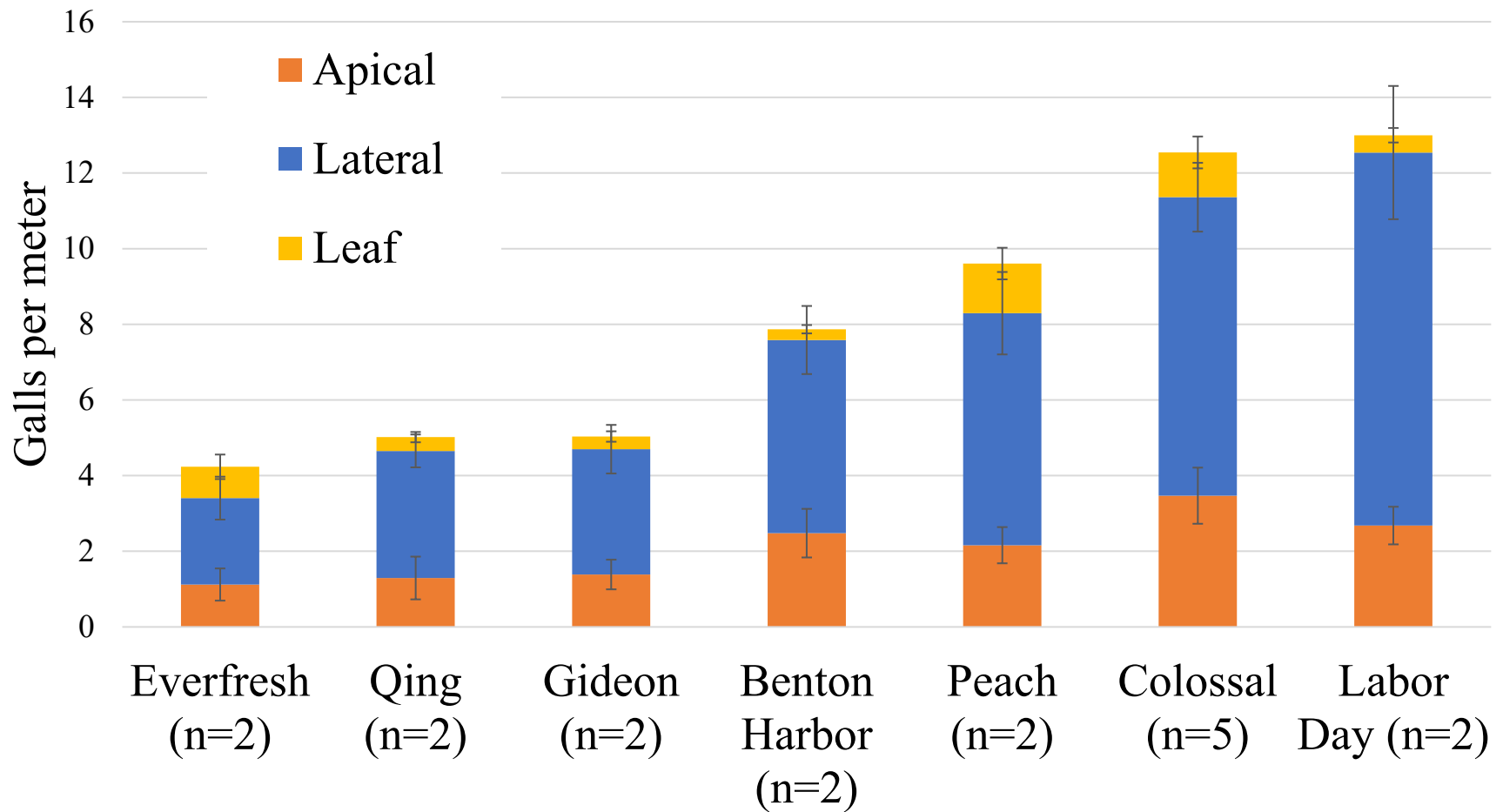


# Average ( $\pm$ SE) gall density (galls per meter) on shoots by cultivar





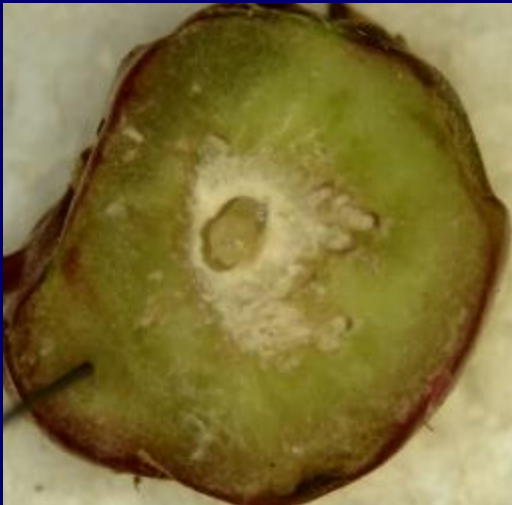
# Average ( $\pm$ SE) density of galls by location & cultivar at SWMREC



# Gall Dissections

- Randomly selected, measured & dissected galls from collected shoots; Total of 476 galls in 2017 & 395 in 2018
- Recorded gall contents:
  - ACGW life stage
  - Live or dead
  - Chamber condition

ACGW larva



ACGW pupae

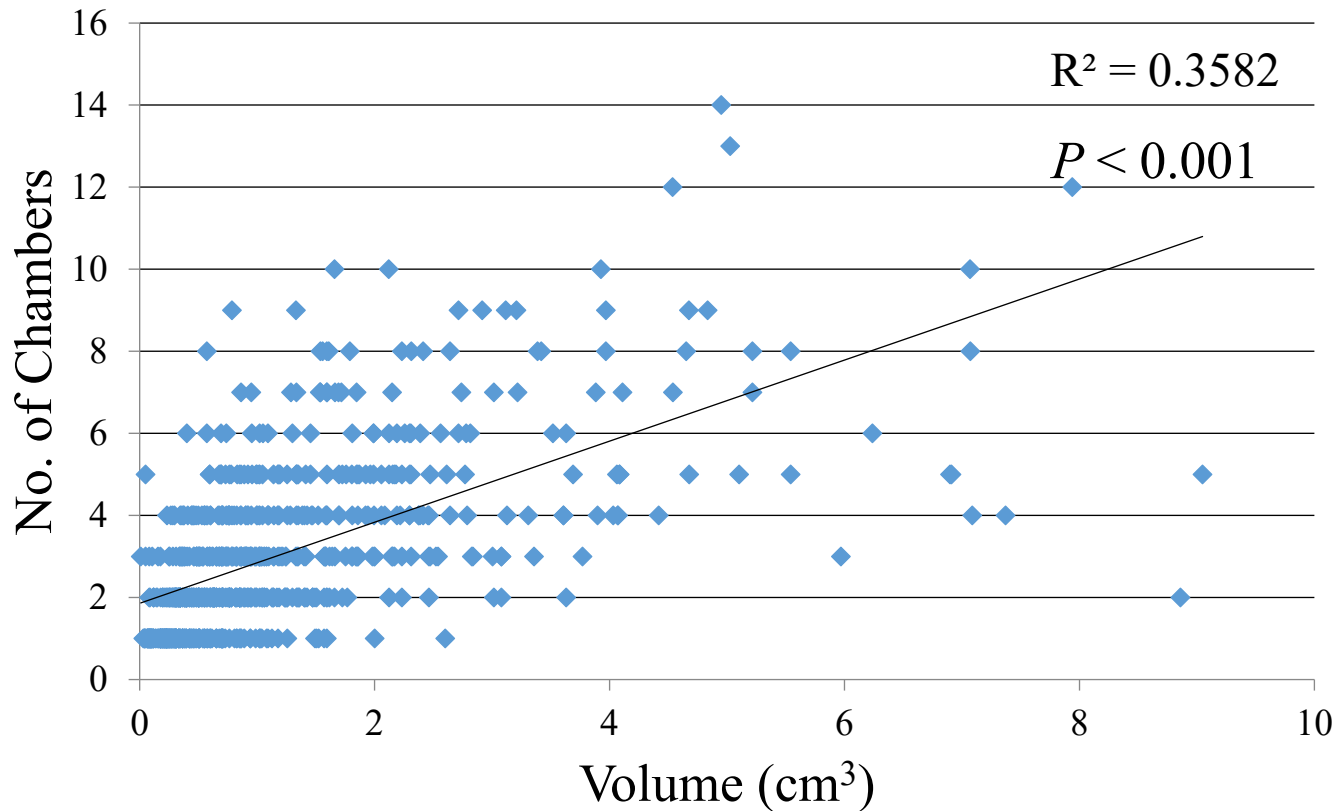


Chamber with fungus



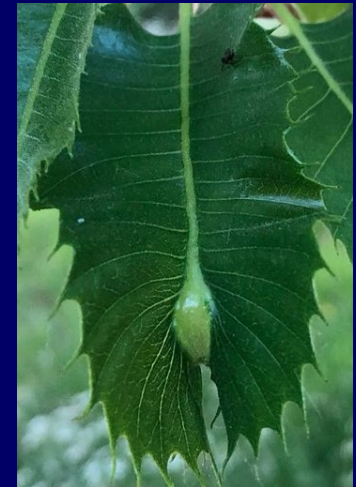
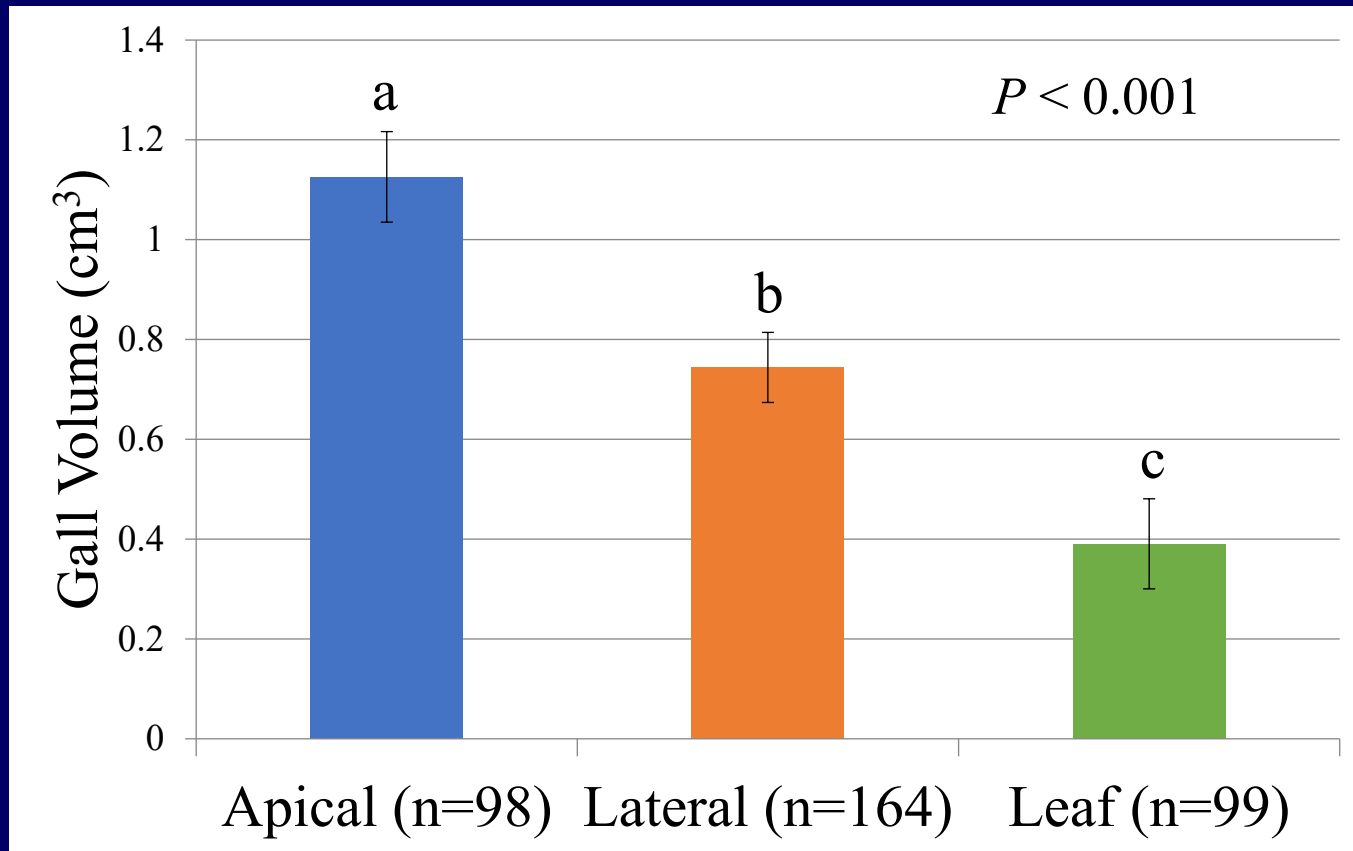
# Gall Volume & Number of Chambers

Galls were measured (834), volume calculated & chambers counted. Most (51%) galls had 1-2 chambers, 39% had 3-5 chambers & 10% of galls had > 5 chambers.

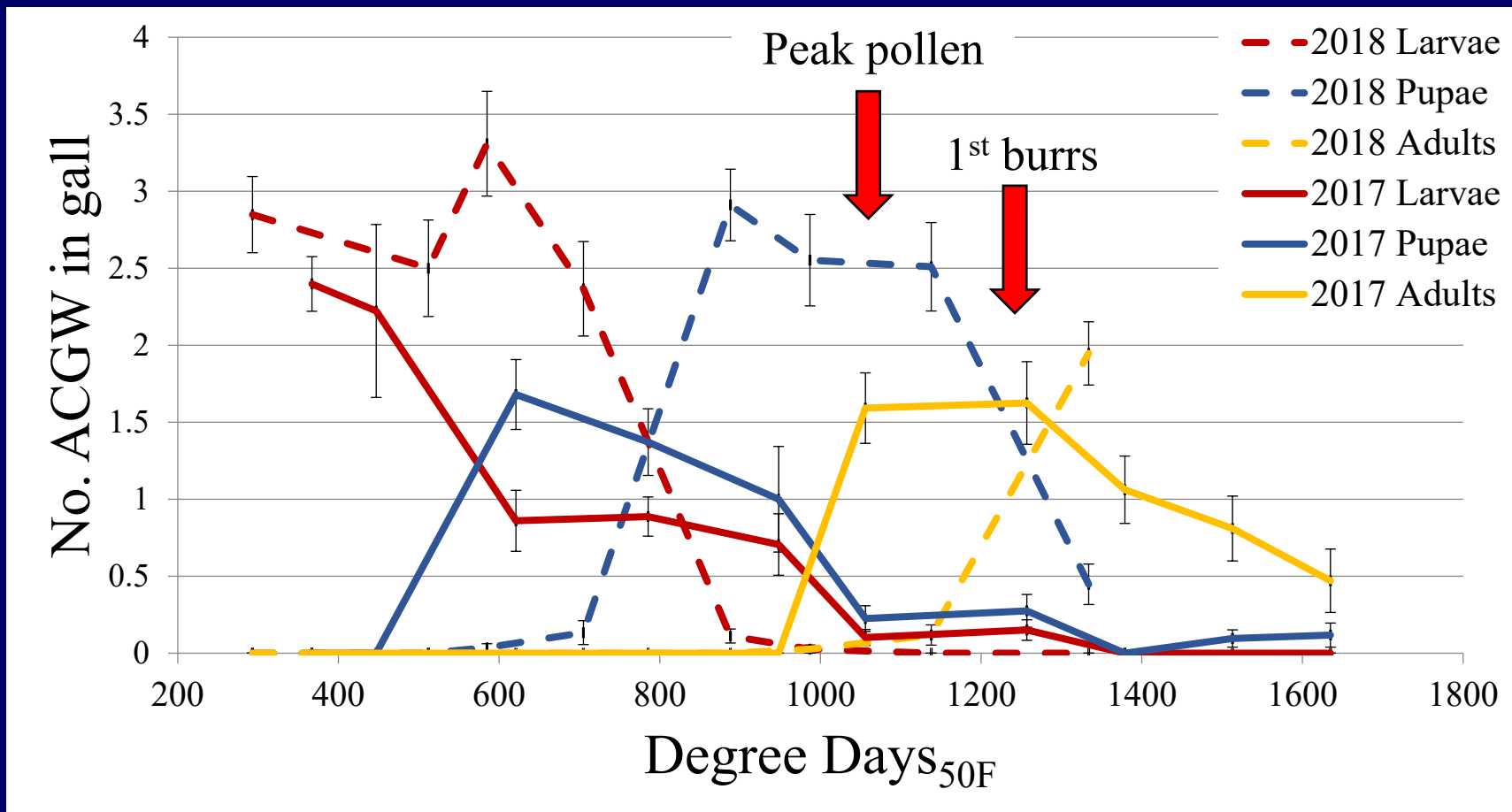


# Gall Volume & Gall Location

Location recorded & volume calculated for 361 galls.  
Volume ranged from 0.009 cm<sup>3</sup> to 8.86 cm<sup>3</sup>. Only 19% of all the galls had volumes over 1.0 cm<sup>3</sup>.



# Average ( $\pm$ SE) number of live ACGW in galls by life stage & cumulative degree days



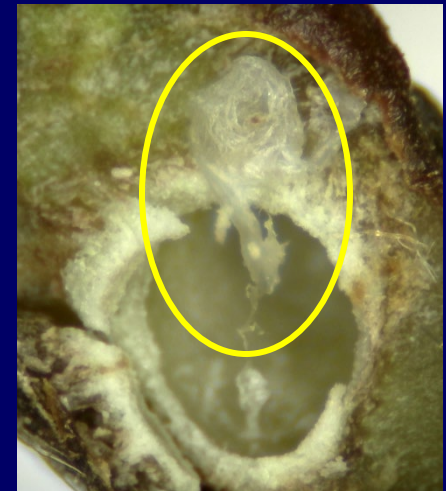
- Larvae: Mid May to early July
- Pupae: Early June to mid July
- Adults: Late June to late July

# ACGW mortality from unknown causes & fungus

| Site | 2017 Death | 2018 Death | 2017 Fungus | 2018 Fungus |
|------|------------|------------|-------------|-------------|
| 1    | 15.8%      | 3.5%       | 1.5%        | 0%          |
| 2    | 13.4%      | 0%         | 3.2%        | 0%          |
| 3    | 6.4%       | 2.7%       | 2.1%        | 0.5%        |
| 4    | 29.7%      |            | 5.5%        |             |
| 5    | 23.1%      | 2.5%       | 7.7%        | 2.5%        |
| 6    |            | 3.5%       |             | 0%          |
| 7    |            | 4.1%       |             | 2.5%        |
| 8    |            | 2.6%       |             | 0%          |



Dead pupa



Dead larva

## Potential Relevance

- Gall dissections provide information about ACGW seasonal development in relation to calendar date & cumulative degree days.
- Assess differences in development among cultivars
- Quantify mortality rates by life stage.



# ACGW Adult Trapping - Methods

Yellow sticky traps were used to capture adult ACGW

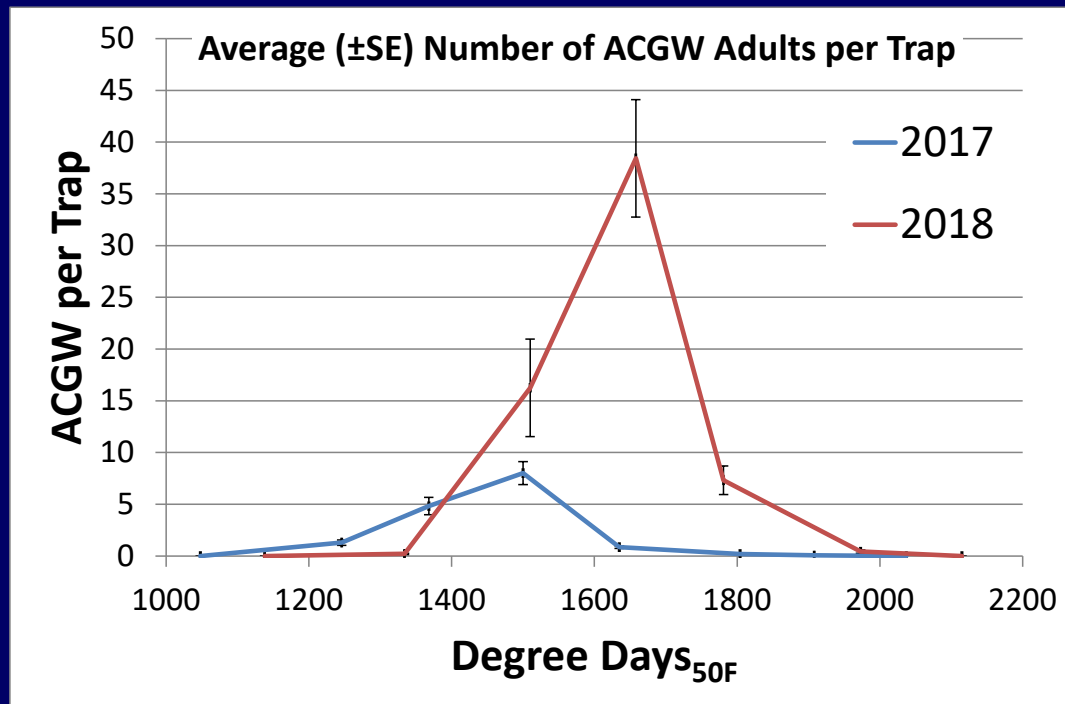
|                       | 2017              | 2018                                      |
|-----------------------|-------------------|---|
| Sticky traps deployed | 21 June to 15 Aug | 2 July to 8 Aug                           |
| Trapped at...         | 11 sites          | 15 sites                                  |
| Traps collected       | Weekly            | Weekly at 7 sites<br>Bi-weekly at 8 sites |





# ACGW Adult Trapping

- Peak emergence in 2017: 19 July
  - 53% between 13 July and 19 July
- Peak emergence in 2018: 24 July
  - 68% between 18 July and 24 July
- Overall more adults captured in 2018
  - Trapped at more fields with multiple years of ACGW

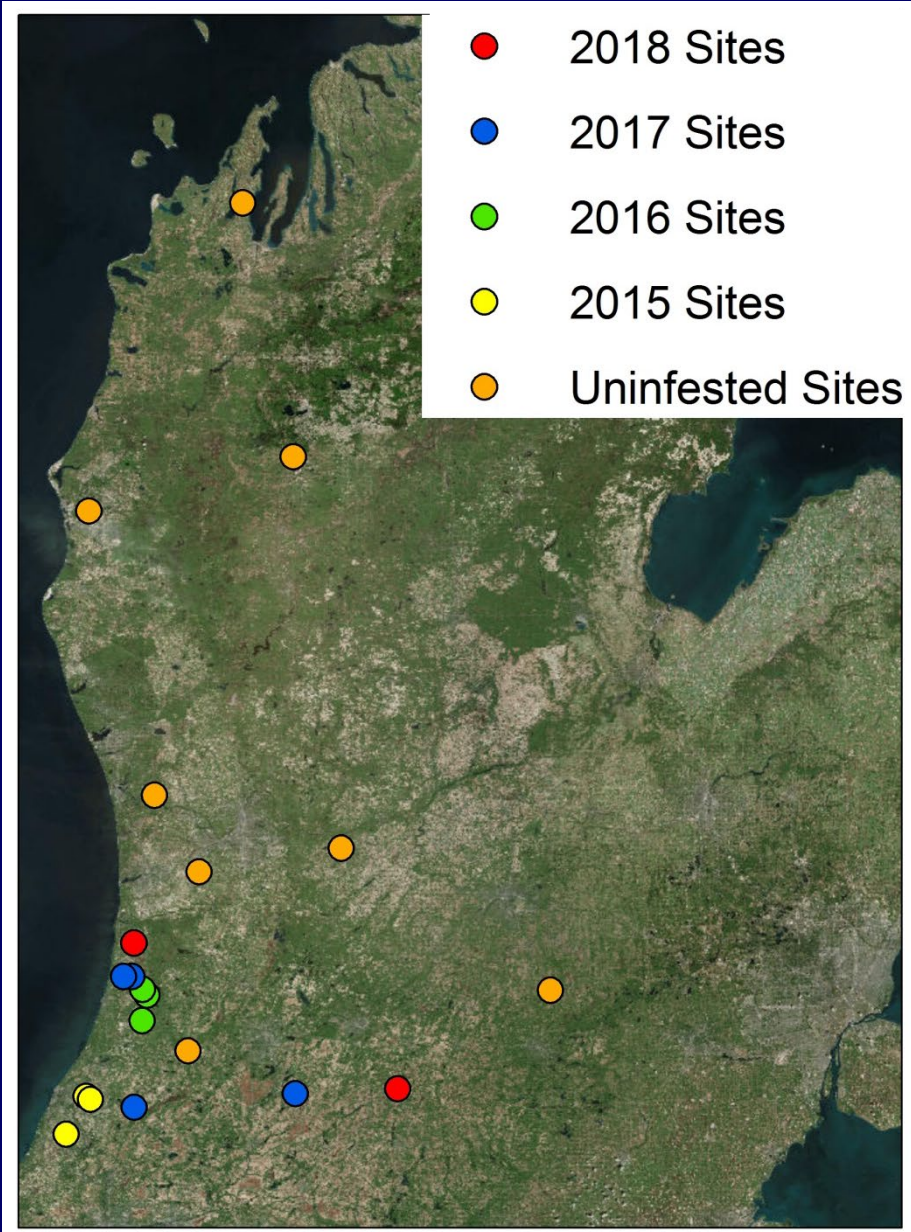


## Potential Relevance

- If control is needed, insecticide cover sprays would be applied when ACGW adults are active.
- 2017: Adult ACGW active from 1250-1900 DD<sub>50F</sub>, corresponding to 6 July to 8 August.
- 2018: Adult ACGW active from 1140-1980 DD<sub>50F</sub>, corresponding to 3 July to 9 August.



# ACGW Annual Spread



- In 2015 and 2016, infested sites were located by observing galls
- In 2017 and 2018, infested sites were located by trapping for adult ACGW

| Year to Year | Max Spread (miles) |
|--------------|--------------------|
| 2015 – 2016  | 26                 |
| 2016 – 2017  | 39                 |
| 2017 – 2018  | 23                 |

# Local abundance & distribution of galls

- Systematically evaluated one site invaded in 2014 & one site invaded in 2015 for 2 years.
- Qualitatively assessed gall abundance on each tree; Rated gall abundance on 0-5 scale.

| Rank | Relative gall abundance                |
|------|--|
| 0    | Absent                                 |
| 1    | $\leq 10$ visible                      |
| 2    | $> 10$ but scattered                   |
| 3    | A few clusters of abundant galls       |
| 4    | Galls abundant in some parts of canopy |
| 5    | Galls abundant throughout canopy       |



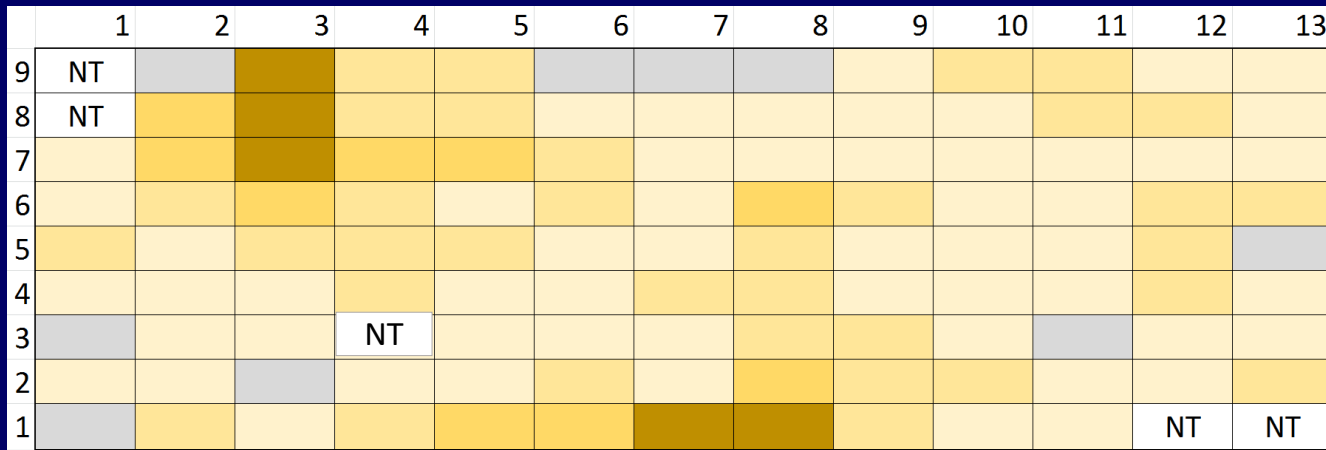
# Spatial Analysis – Semivariogram

- Spatial autocorrelation: index of whether nearby trees influence infestation of a selected tree; can be + or -
- Semivariance: the strength of the relationship
- Semivariogram: how semivariance changes with distance



# Within-field abundance & distribution of trees with galls: Site invaded in 2015

## Site 5 – 2017 evaluation

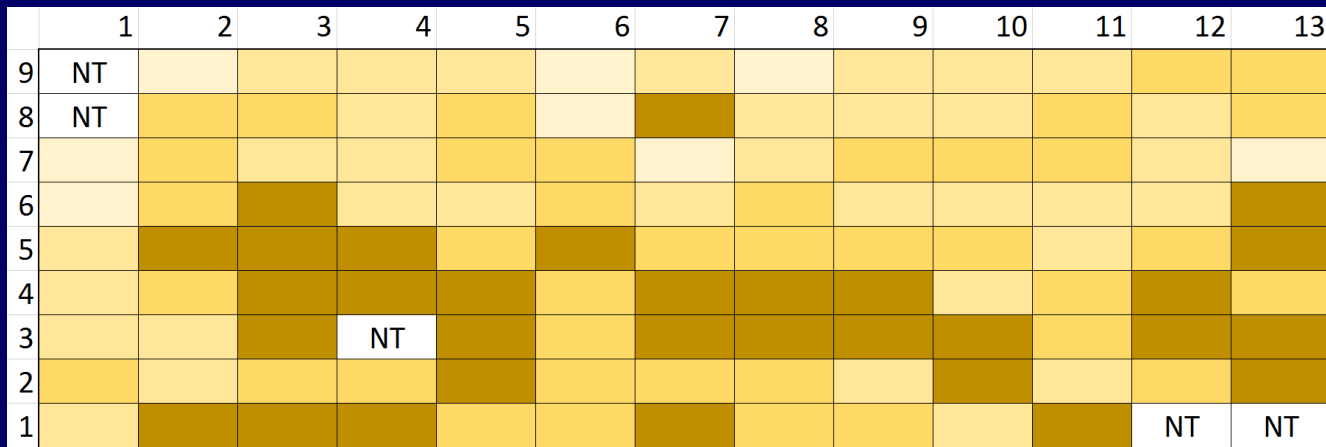


4% of trees  
rated as 4 or 5

### Gall Abundance

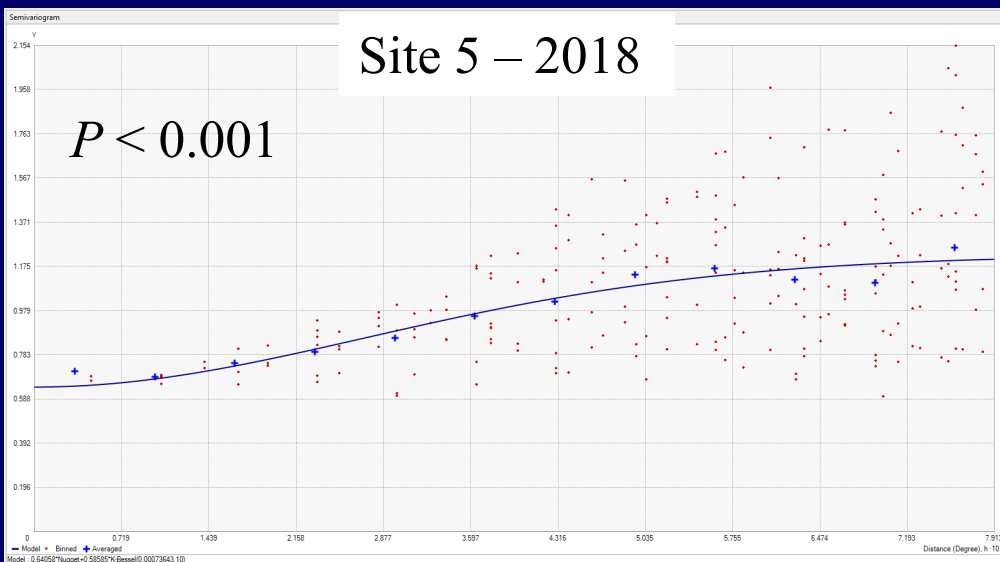
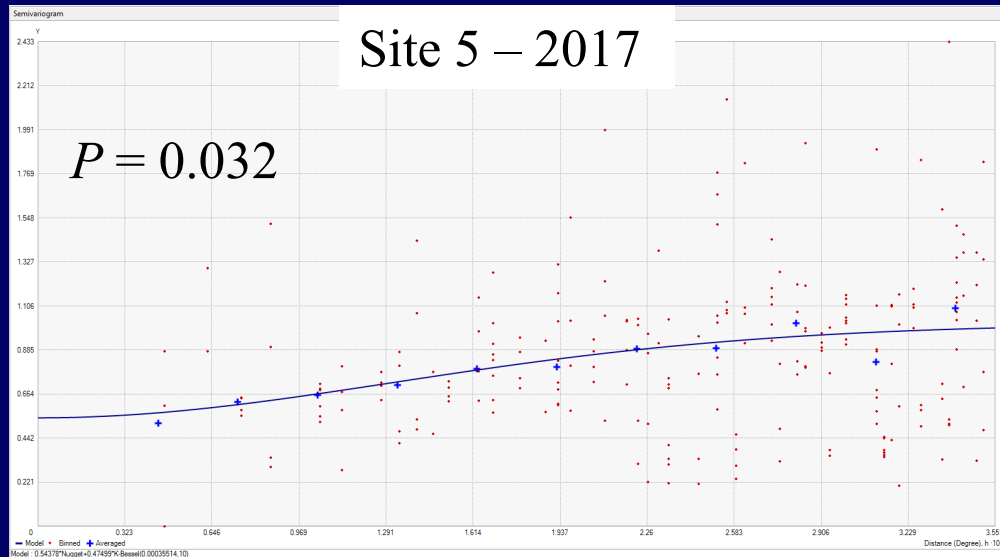


## Site 5 – 2018 evaluation



28% of trees  
rated as 4 or 5

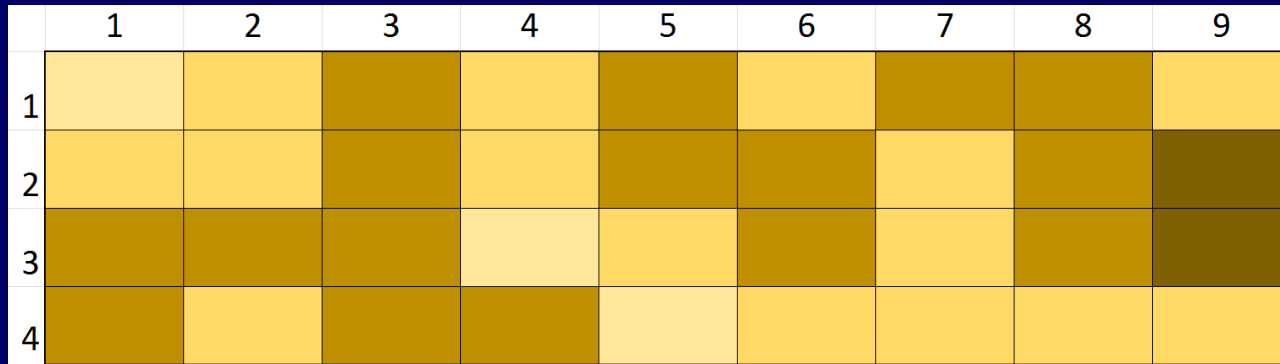
# Semivariograms of Site 5 – 2107 & 2018



- Semivariance increases as distance increases
- Indicates strong spatial dependence
- Trees near infested trees are more likely to be infested & have higher gall densities than trees further away.

# Within-field abundance & distribution of trees with galls: Site invaded in 2014

## Site 2 – 2017 evaluation

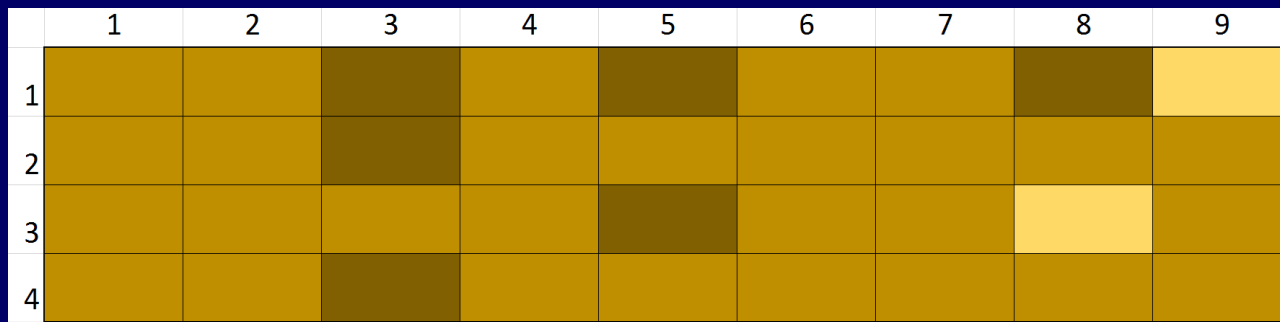


50% of trees  
rated as 4 or 5

### Gall Abundance



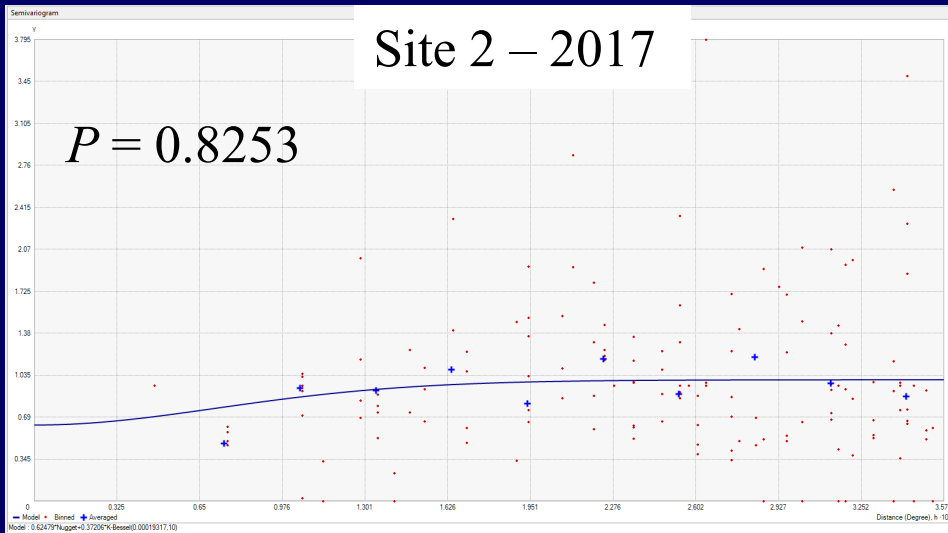
## Site 2 – 2018 evaluation



94% of trees  
rated as 4 or 5

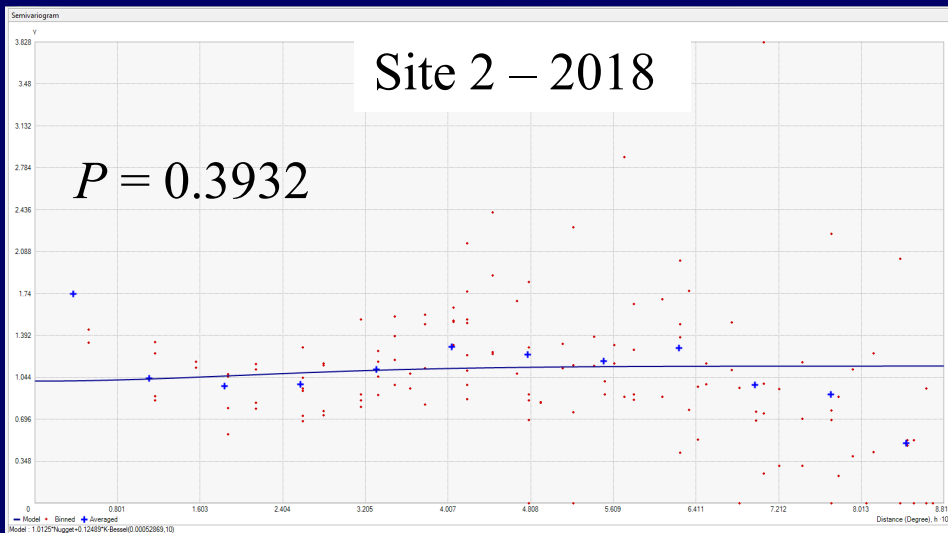


# Semivariograms of Site 2 – 2017 & 2108



➤ Weak relationship between semivariance and distance indicates little or no spatial dependence

➤ This field has a longer infestation history & is smaller than Site 5. All trees have become infested & most have high gall densities. Location of infested & uninfested trees is no longer relevant.



## Potential Relevance

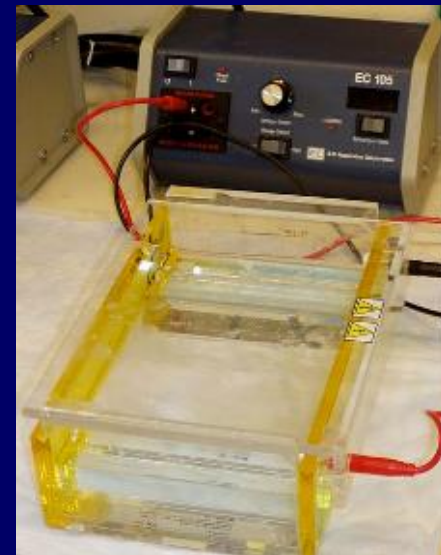
---

- Monitoring gall abundance & distribution provides information on ACGW dynamics & local spread.
- Compare newly invaded fields & fields with a longer history of ACGW
- Project ACGW dispersal & population increase within individual orchards.

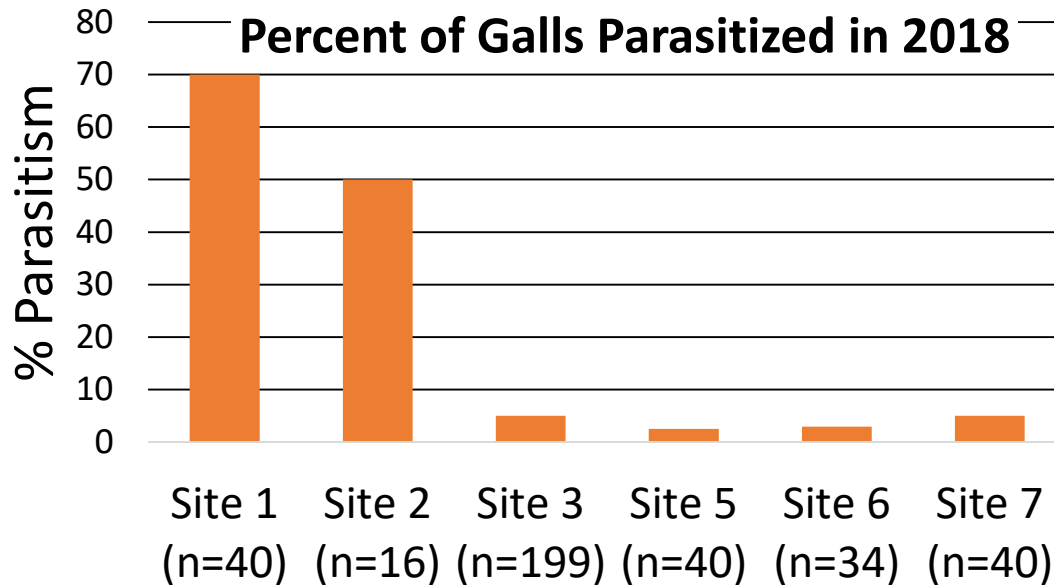
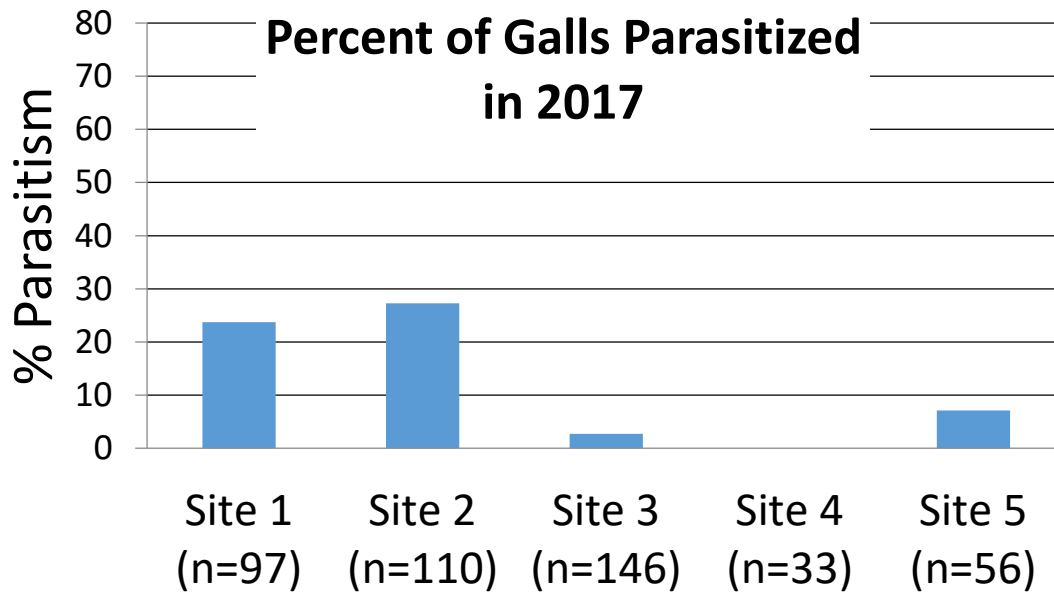


# *Torymus sinensis* parasitism

- *T. sinensis* was not intentionally introduced into Michigan.
- Non-ACGW larvae we suspected to be parasitoids were set aside during gall dissections.
- Extracted DNA from presumed parasitoid larvae; Used PCR to identify species, in cooperation with Dr. Medina Mora.



# Biocontrol of ACGW – Parasitism by Site



## Potential Relevance

- *Torymus sinensis* is established in at least 6 southwest Michigan orchards.
- Overall, an average ( $\pm$  SE) of  $9 \pm 3.6$  % of larvae were killed by this parasitoid; range was 1 to 39% of larvae.
- Parasitoid is spreading; galls with parasitoid larvae were collected 38 miles from sites infested in 2015.
- Mortality from *T. sinensis* may slow ACGW population growth & gall abundance.



# Systemic Insecticide

## ➤ Trunk Injections

- Fall 2017: Imidacloprid
- Spring 2018: Imidacloprid & Ememectin Benzoate
- Fall 2018: Imidacloprid & Ememectin Benzoate

## ➤ Tissue Samples (2018)

- Leaves
- Galls
- Pollen
- Nuts



# Potential Relevance

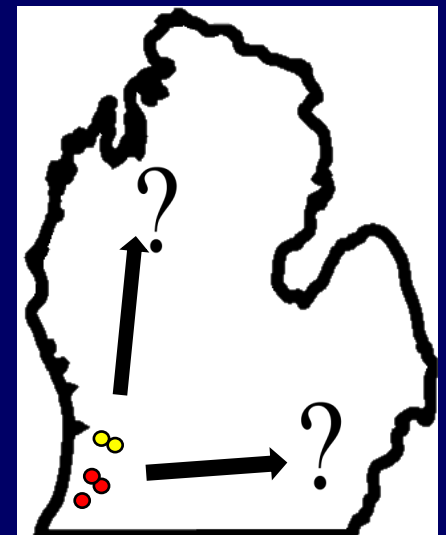
---

- Will trunk injected insecticide control ACGW?
- Translocation & persistence of systemic insecticide in selected tissues of chestnut?
- Broader interest in systemic insecticides & physiology of nut-bearing trees.



# Ongoing work

- Evaluate systemic insecticides as ACGW control method
  - Assess translocation, persistence and residues
  - Imidacloprid and emamectin benzoate
- Spatial analysis
  - Macro and micro spread data
  - 2019 within-field gall abundance evaluation
  - 2019 ACGW adult trapping
- Develop an integrated management strategy for ACGW





# Acknowledgments



Undergraduate Research Assistants – MSU  
Special thanks to the growers who allowed us to access their orchards & sample their trees!



**Funding** provided by MSU Project GREEN, the Michigan Chestnut Growers Association, Rogers Reserve & the Michigan Nut & Fruit Growers Association.





Questions?